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Predicting venous thromboembolism risk from ETP-based nAPCsr assay: toward early assessment of combined oral contraceptives.



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PII: S2475-0379(25)00609-0

DOI: <https://doi.org/10.1016/j.rpth.2025.103285>

Reference: RPTH 103285

To appear in: *Research and Practice in Thrombosis and Haemostasis*

Received Date: 23 October 2025

Accepted Date: 23 November 2025

Please cite this article as: Morimont L, Creinin MD, Gaspard U, Foidart J-M, Douxfils J, Predicting venous thromboembolism risk from ETP-based nAPCsr assay: toward early assessment of combined oral contraceptives., *Research and Practice in Thrombosis and Haemostasis* (2026), doi: <https://doi.org/10.1016/j.rpth.2025.103285>.

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1 **Predicting venous thromboembolism risk from ETP-based nAPCsr assay: toward early**
2 **assessment of combined oral contraceptives.**

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17 **Word counts:** Abstract: 242 ; Text : 2643

18 **Figures:** 5

19 **Tables:** 2

20 **Complementary material:** 1 figure

21 **Conflicts of Interest:**

22 LM: is employee of QUALiblood s.a.

23 MDC: has received speaking honoraria from Gedeon Richter and Mayne, served on Advisory
24 Boards for Estetra SRL, has stock options with Femasys, and has consulted for Gedeon
25 Richter, Mayne, Medicines360, and Merck Sharpe Dohme. The Department of
26 Obstetrics and Gynecology, University of California, Davis, receives contraceptive
27 research funding for Dr. Creinin from Femasys, Organon, Sebela, and Myovant
28 (Sumitomo Pharma).
29 UG: has received consultancy fees from Mithra Pharmaceuticals.
30 JMF: has received speaking honoraria from Gedeon Richter and Mayne, served on Advisory
31 Boards for Estetra SRL, and has consulted for Gedeon Richter.
32 JD: consultancy fees from Gedeon Richter, Estetra, and Mithra Pharmaceuticals. He is also
33 the founder of QUALIblood sa, a company involved in the development and validation
34 of the ETP-based activated protein C resistance assay.

35 **ABSTRACT**

36 **Background:** Combined oral contraceptives (COCs) increase venous thromboembolism (VTE)
 37 risk, depending on estrogen type, dose, and the progestin. While epidemiological studies
 38 provide insight into these risks, they require years to complete. The normalized activated
 39 protein C sensitivity ratio (nAPCsr), a standardized assay of acquired APC resistance, has
 40 emerged as a potential biomarker for COC-induced VTE risk.

41 **Objectives:** To develop a population-based *in silico* model predicting VTE risk associated with
 42 various COC formulations based on their mean nAPCsr values.

43 **Methods:** We analyzed 200 plasma samples from non-COC users and 257 from users of nine
 44 different COCs. We constructed an exponential model correlating mean nAPCsr of five COCs
 45 with their available population-based VTE relative risk extracted from *a published* meta-
 46 analysis. We assessed model performance using R^2 , Spearman's rank correlation, and Root
 47 Mean Square Error (RMSE) and performed a sensitivity analysis by excluding COC non-users.
 48 We then estimated population-based VTE risks for the four COCs not used in model
 49 construction.

50 **Results:** The model demonstrated high predictive accuracy ($R^2=0.96$, RMSE=0.21, Spearman's
 51 $Rs=1$) and remained robust despite group size imbalance. Predicted VTE risks for EE 30 μ g-
 52 dienogest 2mg, EE 20 μ g-drospirenone 3mg, E2 1.5mg-nomegestrol acetate 2.5mg, and E4 15
 53 mg-drospirenone 3mg were 4.36, 3.43, 1.50 and 1.45, respectively, consistent with or
 54 complementary to existing epidemiological evidence.

55 **Conclusions:** Our model based on mean nAPCsr provides a reliable, biomarker-based
 56 approach for predicting population-based COC-related VTE risk. This strategy could help
 57 shorten the time between product launch and population-based risk assessment.

58 **Keywords:** activated protein C resistance; combined oral contraceptives; estrogens; risk
59 assessment; venous thromboembolism

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60 **INTRODUCTION**

61 Combined hormonal contraceptives (CHCs) containing the synthetic estrogen ethinyl
62 estradiol (EE) are associated with an increased venous thromboembolism (VTE) risk compared
63 to non-users. This risk varies from 2.2-6.6 times higher, depending on the EE dosage, the
64 associated progestin and various clinical factors such as body mass index (BMI), diabetes,
65 mellitus, hypertension, or polycystic ovary syndrome.[1-4]

66 Over the past decade, new combined oral contraceptives (COCs) containing body
67 identical estrogens such as estradiol (E2) and estetrol (E4), have been introduced with the aim
68 of significantly reducing VTE risk compared to EE-containing regimens. A meta-analysis of
69 population-based observational studies and a pharmacovigilance analysis using the
70 Eudravigilance database have provided data to support lower VTE risk with these body
71 identical estrogens.[5, 6]

72 In current COC regulatory pathways, surrogate coagulation markers are typically
73 evaluated during safety clinical trials.[7] To date, no surrogate markers have been clearly
74 categorized as directly correlating with COC-related VTE risk. Consequently, our only resources
75 for VTE risk estimations are phase IV population-based post authorization safety studies
76 (PASS) which often take several years to complete. For example, results from the E2-
77 nomegestrol acetate PASS were published in 2021, nearly a decade after initial marketing.[8]

78 COC use is associated with acquired activated protein C resistance (APCr), a
79 dysregulation of hemostasis that occurs independent of factor V (FV) Leiden genetic
80 mutations.[9, 10] Among the several methods developed to detect APCr, endogenous
81 thrombin potential (ETP)-based APCr assay, introduced in 1997, has proven to be the most
82 suitable for assessing COC-induced coagulation changes.[11-15] This method is currently

83 endorsed by European Medicines Agency (EMA) guidelines for the assessment of new
84 steroid contraceptives in women.[7] Moreover, the International Society on Thrombosis and
85 Haemostasis (ISTH) Subcommittee on Plasma Coagulation Inhibitors declared the ETP-based
86 APCr assay as the test of choice for assessing hormone-induced acquired APCr.[16] Prior to
87 2020, this assay suffered from a lack of standardization, hampering study-to-study
88 comparison. Since 2020, when the test underwent analytical validation, it has been available
89 as a fully harmonized measurement scale, known as the normalized activated protein C
90 sensitivity ratio (nAPCsr).[17] Because early evidence suggests a correlation of nAPCsr with
91 COC-associated VTE risk, [12, 18] we aimed to develop a population-based *in silico* model. This
92 model integrates nAPCsr values with epidemiological data on VTE risk related to specific COCs,
93 to predict the VTE risk linked to newly formulated CHC. To accomplish our study goals, we first
94 obtained plasma samples from COC and non-COC users to measure nAPCsr using the validated
95 ETP-based APCr assay [17], and then developed an *in silico* model incorporating the measured
96 nAPCsr values with VTE risk based on epidemiological data.

97 **2.0 MATERIAL & METHODS**

98 **2.1 Sample collection**

99 We collected plasma samples from 2 different sources: a phase II clinical trial (NCT
100 02957630) conducted in 2016-2017 for which sample collection have been previously
101 described [19], and the NAmur Biobank-eXchange (NAB-X), the registered biobank
102 (notification number BB190116) affiliated with the University of Namur (Namur, Belgium). We
103 obtained these latter specimens from 18 different blood donation campaigns organized since
104 2018 at the University of Namur (Namur, Belgium) that recruited volunteers to donate blood
105 for research purposes, in accordance with approval from the Ethical Committee of the Centre

106 Hospitalier Universitaire, Université Catholique de Louvain (CHU UCL, Namur, Belgium)
107 (approval number B03920096633). Eligibility criteria for blood donation excluded current
108 pregnancy, individuals with a history of thrombotic or hemorrhagic events, and using
109 antiplatelet or anticoagulant medication or any other drugs known to affect platelet function
110 or coagulation. Study staff obtained written informed consent from participants prior to
111 sample collection and recorded demographic data, medical history, and current medication
112 use. Blood samples were collected in sodium citrate tubes. The first tube, used as a primer,
113 was reserved for genotyping of FV Leiden and prothrombin G20210A mutations using a CE-
114 marked in-vitro diagnostic (IVD) technique (LAMP Human FII&FVL duplex kit, Lacar, Belgium).
115 The remaining tubes were processed to obtain plasma which was then frozen in liquid nitrogen
116 and stored at $\leq -70^{\circ}\text{C}$ in the NAB-X biobank.

117 For the present study, we requested NAB-X plasma samples obtained from women of
118 childbearing age (18-40 years), not carrying a FV Leiden or G20210A prothrombin mutation,
119 not using any hormonal contraceptive method or had been using the same COC for at least 3
120 consecutive cycles, and stored for less than 3 years (to ensure plasma integrity for the ETP-
121 based APC resistance assay, which remains stable for up to 36 months). All samples, whether
122 originating from the clinical trial or from NAB-X, were analyzed within three years after
123 collection. The validated, standardized, and ISO/IEC 17025-accredited method ensures well-
124 controlled batch transitions, thereby maintaining consistent comparability between historical
125 and recent results.

126 **2.2 ETP-based APC resistance assay**

127 The ETP-based APC resistance assay was performed as summarized in **Figure 1**. Briefly, this
128 test is a variant of the thrombin generation assay, a global coagulation test, enabling a

129 continuous overview of clotting over time in a test cuvette. Thrombin generation is measured
130 in the presence and absence of a defined amount of exogenous APC. This amount is batch-
131 specific and determined using a standardized procedure calibrated to achieve 90% inhibition
132 of ETP (i.e. area under the thrombin generation curve) in a healthy pooled plasma.[17] In the
133 absence of APC, the resulting thrombin generation curve reflects all the pro- and anticoagulant
134 reactions that regulate thrombin formation and inhibition. In the presence of exogenous APC,
135 thrombin generation is significantly decreased in a normal plasma sample allowing
136 measurement of the sample sensitivity towards APC. Results are expressed as a ratio, the
137 nAPCsr.

138 **2.3 Statistical analysis**

139 We performed all statistical analyses using GraphPad Prism version 10.6.1 for macOS
140 (GraphPad Software, San Diego, CA, USA, www.graphpad.com). We used a convenience
141 sampling, including all available samples, and stratified them according to the COC used.

142 To assess subgroup homogeneity, we compared age and BMI using Kruskal-Wallis
143 followed by Dunn's multiple comparison, given the non-normal distribution of the data. For
144 nAPCsr values, although the data were normally distributed (with the exception of the non-
145 user group which has a sufficiently large sample size), we used Welch's ANOVA because of
146 unequal variance across subgroups. We identified different clusters based on previously
147 observed phenotypic differences.[18] In this confirmatory context, we applied no correction
148 for multiple comparisons (i.e., unpaired t with Welch's correction) to preserve statistical
149 power and reported unadjusted p-values to reflect the *a priori*-defined comparisons of
150 interest.

151 We created the model using nAPCsr values from COC non-users and COC users of
152 products with known population-based VTE risks, as summarized in the network meta-analysis
153 of *de Bastos et al.*[1] Because we used a convenience sampling strategy reflecting local COC
154 prescribing patterns, we did not include all COCs reported in that meta-analysis, but focused
155 only on those available in our cohort: EE 20 μ g with levonorgestrel 100 μ g, EE 30 μ g with
156 levonorgestrel 150 μ g, EE 20 μ g with desogestrel 150 μ g, EE 30 μ g with desogestrel 150 μ g and
157 EE 35 μ g with cyproterone acetate 2mg. Conversely, no samples were available for the
158 following formulations: EE 50 μ g with levonorgestrel, EE 20 μ g with gestodene, EE 30 μ g with
159 gestodene, EE 35 μ g with norgestimate and EE 30 μ g with drospirenone. We plotted the mean
160 nAPCsr values on the x-axis and the corresponding VTE relative risk (RR) associated with these
161 COCs (compared to non-users) on the y-axis.

162 Using these data, an exponential growth model of the form $y = Y_0 \cdot \exp(k \cdot x)$ was
163 constructed to describe the relationship between nAPCsr values and VTE risk. Considering
164 disparities across subgroups, the model incorporated samples size (N) through weighted
165 fitting. We performed a sensitivity analysis to assess the potential impact of the large
166 reference group (i.e., COC non-users), which was excluded from the curve fitting while
167 preserving the model's anchoring at Y_0 . We assessed model fit using the coefficient of
168 determination (R^2) while the strength of association between nAPCsr and VTE risk was
169 evaluated using Spearman's rank correlation coefficient (Rs).

170 We validated the model using an actual versus predictive plot, in which we plotted
171 predicted VTE risks from the *in silico* simulation against observed risks from *de Bastos* meta-
172 analysis. We assessed quality of fit by visual alignment with the identity line ($y = x$) and
173 quantified using the Root Mean Square Error (RMSE).

174 Finally, to further assess the model's predictive capabilities, we estimated VTE risk for
 175 COC formulations not included in the base model, using their corresponding mean nAPCsr
 176 values: EE 30 μ g–dienogest 2mg, EE 20 μ g– drospirenone 3mg, E2 1.5mg–norgestrel acetate
 177 2.5mg, and E4 15mg–drospirenone 3mg. For the first three products, the predictive values
 178 were compared to existing population-based estimates of VTE risk. For the fourth product, no
 179 epidemiological data were available to enable such comparison.

180 **3.0 RESULTS**

181 **3.1 Clinical data**

182 Our data set included 457 total samples, with 200 from COC non-users and 257 from COC
 183 users. The number of samples for each COC type and limited demographic characteristics (age,
 184 BMI, plasma collection source) for the participant populations are described in **Table 1**. Age
 185 differed between groups ($p<0.001$) but not BMI ($p=0.45$).

186 **Figure 2A** illustrates the distribution of nAPCsr values across subgroups, and
 187 corresponding mean values \pm 95% confidence intervals (CI) are summarized in **Table 2**.
 188 Bioidentical estrogen COCs (i.e., E2 and E4-based COCs) and EE-levonorgestrel COCs showed
 189 the lowest nAPCsr values, with only the bioidentical estrogen COCs displaying a mean nAPCsr
 190 below 3.0. Four distinct clusters of COC-related VTE risk were considered (**Figure 2B**), ranked
 191 in ascending order of mean nAPCsr: (i) COCs containing bioidentical estrogens (ii) EE-
 192 levonorgestrel COCs (iii) EE20-desogestrel or drospirenone COCs and (iv) COC containing EE30-
 193 desogestrel, cyproterone acetate or dienogest ($p <0.005$ for all pairwise comparisons).

194 **3.2 In-silico modeling**

195 **Figure 3** illustrates the *in silico* model developed using nAPCsr from 346 plasma
 196 samples divided into COC non-users (reference group) and five COC subgroups. The remaining

197 111 samples from 4 additional COC groups not included in the initial model, were used to
 198 assess predictive performance.

199 This approach yielded an exponential growth equation: $y=0.6108*exp(0.3886*x)$. The
 200 coefficient of determination (R^2) was 0.9591 and the Spearman correlation coefficient (Rs) was
 201 1, indicating both a strong goodness of fit and a perfect monotonic association between
 202 nAPCsr values and VTE risk. In the sensitivity analysis excluding COC non-users, the
 203 exponential curve was constrained to pass through the reference point ($y_0 = 0.6108$) to
 204 preserve the epidemiological anchoring of the model. The resulting relationship remained
 205 virtually unchanged (**Supplementary Figure 1**), indicating the model's goodness of fit was not
 206 overly dependent on the reference group.

207 Model validation, as shown by the actual versus predicted plot in **Figure 4**, yielded a
 208 RMSE of 0.2029 indicating that, on average, the predicted relative risk varies by 0.2 units from
 209 the actual relative risk. The resulting *in silico* model estimated VTE risks of 4.36 (95%CI 4.29-
 210 4.43) for EE 30 μ g-dienogest 2mg, 3.43 (95%CI 3.39-3.47) for EE 20 μ g-drospirenone 3mg, 1.50
 211 (95%CI 1.47-1.52) for E2 1.5mg-nomegestrol acetate 2.5mg and 1.45 (95%CI 1.43-1.48) for E4
 212 15mg-drospirenone 3mg versus COC non-users (**Figure 5; Table 2**).

213 4.0 DISCUSSION

214 We successfully developed an *in silico* model to estimate VTE risk associated with new CHC
 215 formulations, using nAPCsr measurements in plasma samples. The high weighted coefficient
 216 of determination ($R^2=0.96$) demonstrates that the model accurately reproduces observed VTE
 217 risks trends based on mean nAPCsr values, showing strong consistency with epidemiological
 218 data. The perfect Spearman correlation ($Rs=1$) further supports the model's robustness,
 219 confirming that it preserves the expected hierarchy of VTE risk across COC types. The strong

220 agreement between predicted and observed relative risks (RMSE = 0.20; alignment with the
 221 identity line) confirms the reliability of this computational approach. Sensitivity analyses
 222 excluding non-users further demonstrated that the association between nAPCsr and VTE risk
 223 remains stable across COC groups and is not solely driven by the reference population. The
 224 concordance of the model with available population-based VTE risk estimates [2, 8, 20] and
 225 clinical data [20] suggests that our model may serve as a valuable tool to estimate the VTE
 226 relative risk of COCs for which current epidemiological data are insufficient or lacking.

227 When interpreting the nAPCsr values obtained in this study, several biological and
 228 methodological aspects should be considered. Although an age-related increase in thrombin
 229 generation has been reported [21], this effect is attenuated when expressed as the nAPCsr,
 230 since the latter is defined as a ratio between two thrombin generation conditions (i.e. ETP in
 231 the presence and absence of exogenous APC). Consequently, the significant age-related
 232 differences observed between subgroups are unlikely to have influenced the nAPCsr results.
 233 BMI was evenly distributed across subgroups, thereby minimizing potential bias. However, as
 234 higher BMI is known to influence coagulation factors and increase the risk of venous
 235 thrombosis [22], its impact on the nAPCsr cannot be fully excluded, even though no
 236 association was observed in our study. Finally, smoking, although a well-known cardiovascular
 237 risk factor, predominantly affects the arterial side of thrombosis [23] and is therefore not
 238 expected to influence the nAPCsr, which reflects venous hemostatic mechanisms.

239 Although the increased nAPCsr variability observed within COC subgroups compared
 240 with non-users may partly stem from smaller subgroup sizes, it is more likely attributable to a
 241 methodological factor. Because the relationship between APC concentration and ETP%
 242 inhibition is curvilinear, variability remains low near 90% inhibition (non-users) but increases
 243 to around 50% inhibition (COC-users) where the slope of the curve is steeper.

244 Consistent with previous findings, we confirmed that APC resistance levels depend on
245 both the dosage and the specific nature of the estrogen and the associated progestin [13]. For
246 EE-containing COCs, APC resistance increased with higher EE doses (30–35 µg), particularly
247 when combined with weakly androgenic (e.g. desogestrel), or anti-androgenic progestins (e.g.
248 dienogest and cyproterone acetate) having a neutral profile on the liver. In contrast, for EE-
249 levonorgestrel combinations, the impact of EE dosage on APC resistance appears limited, likely
250 due to a compensatory increase in the levonorgestrel dose, possibly mediated by its
251 antiestrogenic activity.[13] Importantly, COCs body identical estrogens such as E2 or E4
252 demonstrated the lowest degree of APC resistance, with a mean nAPCsr below 3.0, the upper
253 limit of the 95% CI for COC non-users. These results are consistent with existing evidence
254 regarding the safest hemostatic profile of natural estrogen-containing COCs compared to
255 conventional COCs. [19, 24]

256 Although more recent data on COC-associated VTE risk have been published (*Yonis et*
257 *al*, *JAMA* 2025 [25]), we relied on the *de Bastos* meta-analysis for several reasons. First, as a
258 network meta-analysis of multiple studies, this study offers greater external validity and a
259 broader representation of contraceptive users. Second their risk estimates are consistent with
260 the historical literature and regulatory benchmarks, facilitating comparison. Third,
261 methodological differences in the recent cohort study, such as exposure definitions and
262 population characteristics, may explain the discrepancies in rate ratios and complicate their
263 direct integration into our modeling framework. By contrast, conducting an updated network
264 meta-analysis integrating newly available epidemiological data for E2-based COCs represent a
265 valuable next step.

266 A limitation of our analysis is the small number of women evaluated in some
267 subgroups. However, given that the model relies on weighted subgroup means rather than

268 individual values, and that the sensitivity analysis demonstrated its robustness despite sample
269 size disparities, the use of smaller groups appears acceptable for the purpose of this initial
270 modeling approach. Continued evaluation of these outcomes with larger populations would
271 be important to confirm these results.

272 To conclude, our population-based *in silico* model demonstrates reassuring
273 robustness, supporting the reliability of this computational approach used for estimating VTE
274 risk based on nAPCsr values. Ultimately, such modeling could serve a regulatory purpose by
275 allowing earlier risk assessment of new CHCs, without the need to wait nearly a decade for
276 post-marketing epidemiological data before categorizing them.

277 If a biomarker such as the nAPCsr can be assessed during phase II or III clinical trials, it
278 may provide an early indication of VTE risk for new hormonal contraceptives, pending
279 confirmation in phase IV studies. This approach has the potential to save both time and
280 resources for regulatory authorities, pharmaceutical companies, clinicians and patients by
281 delivering timely and reliable VTE risk data that could facilitate the early adoption of
282 innovative hormonal therapies.

283 **FUNDING**

284 This study was financed by the Walloon Region, Belgium and the Federation Wallonie
285 Bruxelles, Belgium.

286 **AUTHOR CONTRIBUTION**

287 LM and JD designed the study. LM and JD analyzed and interpreted the data. LM performed
288 the statistical analyses and wrote the original draft. JD, MDC, JMF and UG provided input and
289 critical review of the manuscript. All authors revised and approved the final version.

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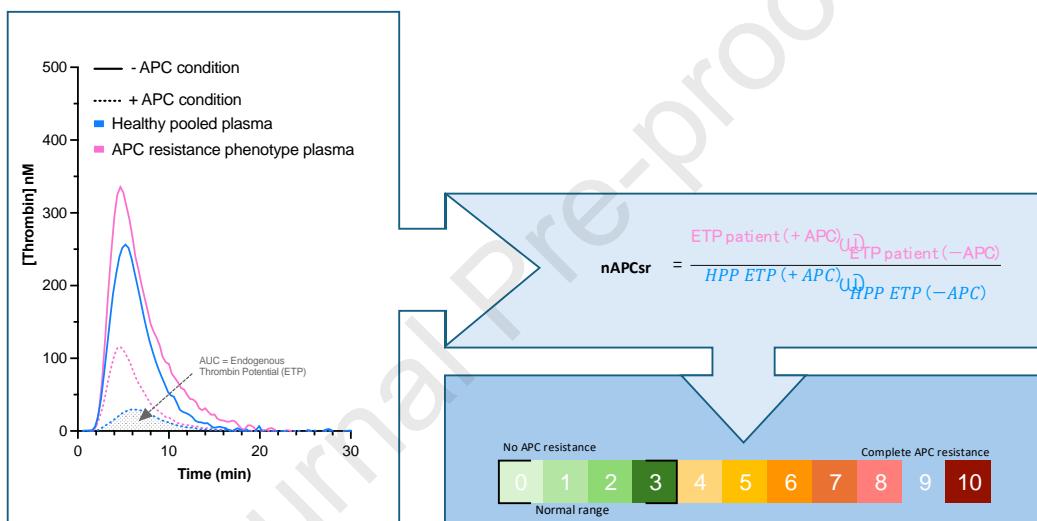
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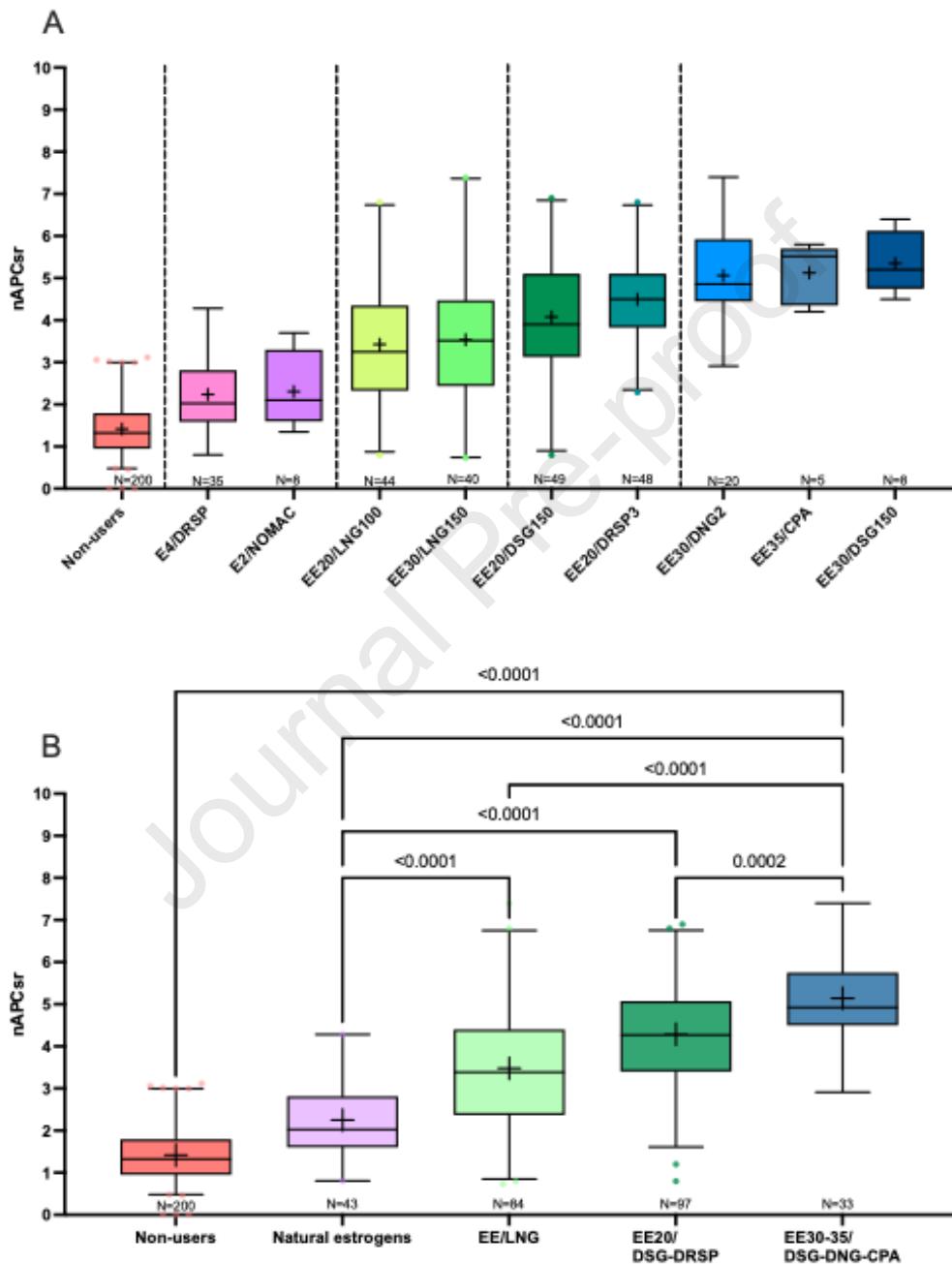
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Figure 1: Thrombin generation curves in absence (continuous line) and in presence of activated protein C (APC) (dotted lines) of a healthy pooled plasma (blue) and an APC resistant phenotype plasma (pink) along with the normalized APC sensitivity ratio (nAPCsr) scale. The reference plasma used to calculate nAPCsr values is derived from pooled plasma of healthy individuals (men and women (in a 1:1 ratio) not using hormonal contraception, not carrier of FV Leiden or prothrombin G20210A mutations). In presence of APC, the ETP (i.e., area under the curve) decreases by 90%, corresponding to a nAPCsr value of 1. In contrast, the APC resistant phenotype plasma, typically seen in women using ethinylestradiol-containing products, shows increased thrombin generation both with and without APC, when compared to the reference plasma. On the nAPCsr scale, this results in a value above the upper limit of the normal range.



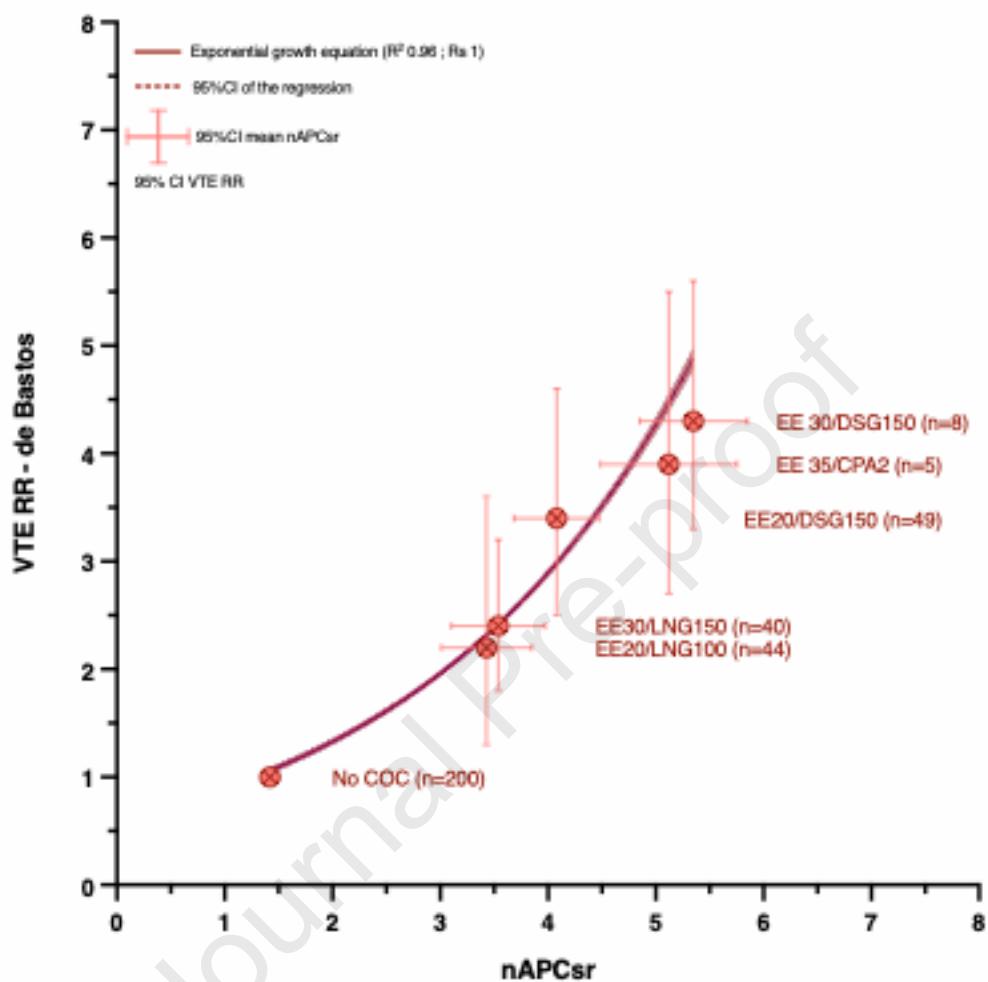
Abbreviations: APC, activated protein C; AUC, area under the curve; ETP, endogenous thrombin potential; HPP, healthy pooled plasma; nAPCsr, normalized activated protein C sensitivity ratio

Figure 2: Box-and-Whisker plot of nAPCsr across each study subgroups (A) and identified clusters (B). The box represents the central 50% of the data, with the lower edge indicating the 1st quartile and the upper edge the 3rd quartile. The line inside the box corresponds to the median while the cross indicates the mean. The whiskers extend to the 2.5th and 97.5th percentiles. Differences between clusters were assessed by an analysis of variance with unpaired t test with Welch's correction. Threshold for significance was set at 0.05.



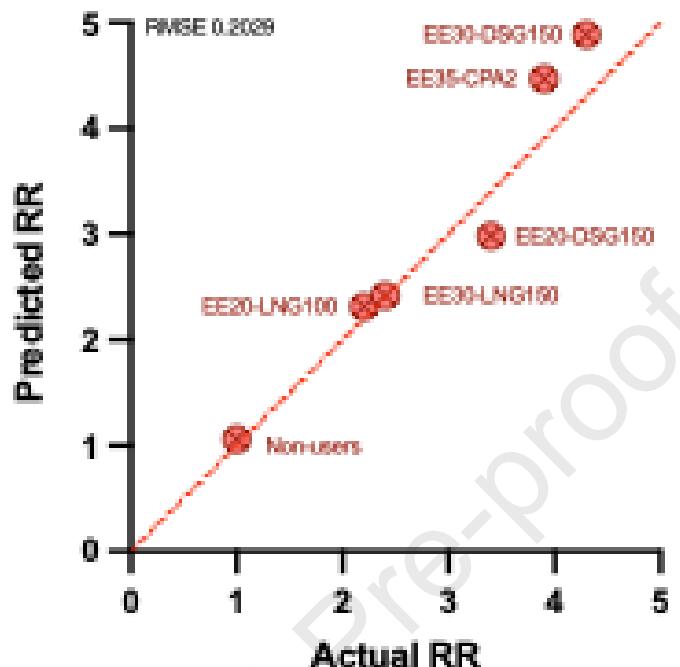
Abbreviations: COC, combined oral contraceptive; CPA, cyproterone acetate; DNG, dienogest; DRSP, drospirenone; DSG, desogestrel; EE, ethinylestradiol; E2, estradiol; E4, estetrol; LNG, levonorgestrel; nAPCsr, normalized activated protein C sensitivity ratio; NOMAC, nomegestrol acetate.

Figure 3: *In silico* modeling based on nAPCsr data and venous thromboembolism relative risk estimates as reported in the meta-analysis of *de Bastos et al.*



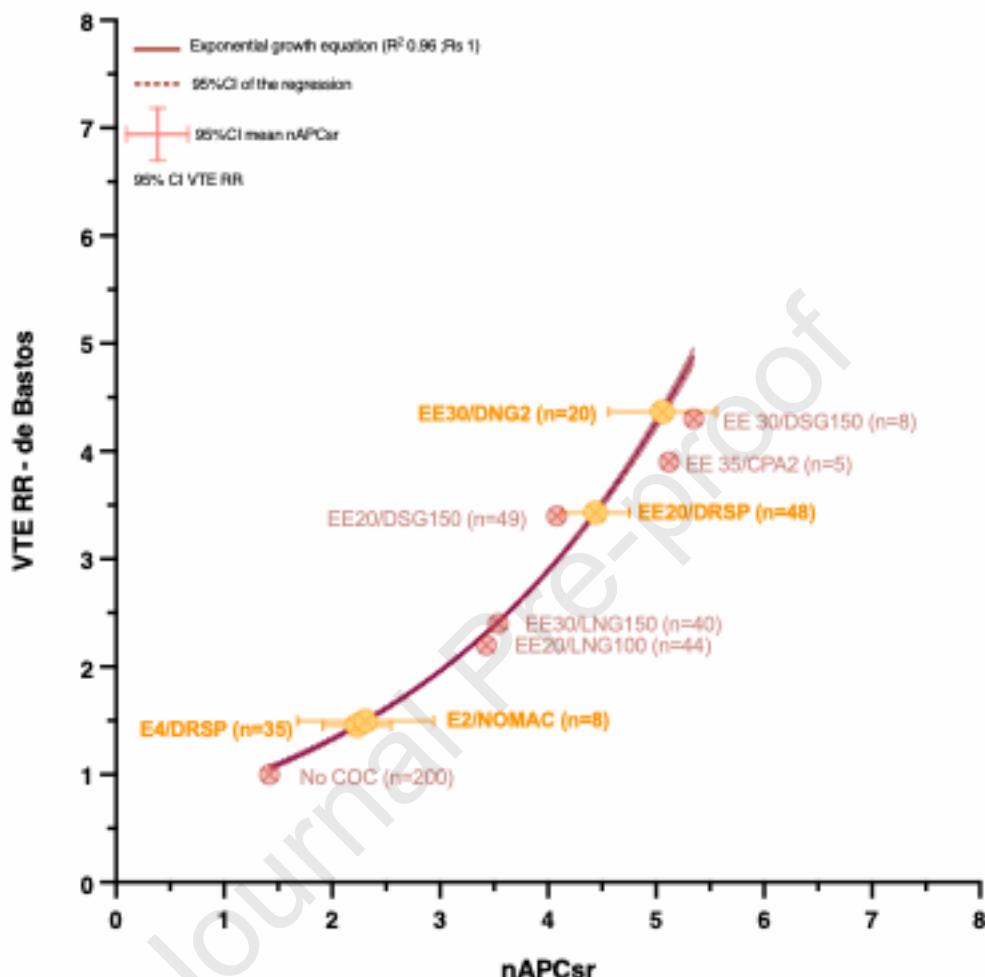
Abbreviations: CI, confidence interval; COC, combined oral contraceptive; CPA, cyproterone acetate; DRSP, drospirenone; DSG, desogestrel; EE, ethinylestradiol; LNG, levonorgestrel; nAPCsr, normalized activated protein C sensitivity ratio; RR, relative risk; SD, standard deviation; VTE, venous thromboembolism

Figure 4: Predicted versus actual relative risk of venous thromboembolism based on nAPCsr data. Actual relative risk values are taken from the meta-analysis of *de Bastos et al.* The dashed line represents the identity line, indicating perfect agreement between predicted and observed values.



Abbreviations: CPA, cyproterone acetate; DRSP, drospirenone; DSG, desogestrel; EE, ethinylestradiol; LNG, levonorgestrel; nAPCsr, normalized activated protein C sensitivity ratio; RR, relative risk; RMSE, Root Mean Square Error.

Figure 5: Venous thromboembolism risk estimates for four combined oral contraceptives based on our *in silico* modeling: estetrol (E4) 15mg with drospirenone 3mg, estradiol (E2) 1.5mg with nomegestrol acetate 2.5mg, ethinylestradiol (EE) 20 μ g with drospirenone 3mg, and ethinylestradiol (EE) 30 μ g with dienogest 2mg.



Abbreviations: *Cl*, confidence interval; *COC*, combined oral contraceptive; *CPA*, cyproterone acetate; *DRSP*, drospirenone; *DSG*, desogestrel; *EE*, ethinylestradiol; *E2*, estradiol; *E4*, estetrol; *GSD*, gestodene; *LNG*, levonorgestrel; *nAPCsr*, normalized activated protein C sensitivity ratio; *NOMAC*, nomegestrol acetate; *RR*, relative risk; *SD*, standard deviation; *VTE*, venous thromboembolism

Table 1. Demographic data of participants that provided samples for endogenous thrombin potential-based nAPCr assay

	TOTAL	Non-COC users	EE 20µg - LNG 100µg	EE 30µg - LNG 150µg	EE 20µg - DSG 150µg	EE 30µg - DSG 150µg	EE 35µg - CPA 2mg	E4 15mg - DRSP 3mg	E2 1.5mg - NomAc 2.5mg	EE 20µg - DRSP 3mg	EE 30µg - DNG 2mg	P-value*
Number	409	200	44	40	49	8	5	35	8	48	20	
Age (years)												
Mean (±SD)	23 (±5)	24 (±6)	21 (±2)	24 (±5)	21 (±3)	22 (±2)	20 (±2)	26 (±6)	22 (±2)	24 (±5)	22 (±2)	<0.001
Range	18-47	18-47	18-28	18-44	18-29	19-26	18-22	19-43	20-25	18-40	19-27	
BMI (kg/m²)												
Mean (±SD)	22.6 (±3.2)	22.7 (±3.4)	22.3 (±3.2)	22.4 (±3.2)	22.3 (±2.6)	21.6 (±2.2)	22.3 (±2.2)	23.3 (±2.9)	23.1 (±3.6)	21.8 (±2.4)	21.6 (±2.7)	0.45
Range	16.9-38.9	16.9-38.9	17.6-28.7	17.4-29.8	17.7-31.3	18.3-24.8	20.2-25.8	19.0-30.0	19.5-30.0	18.4-28.7	18.5-29.8	
Collection Source												
Biobank	251(62%)	104(52%)	44(100%)	12(30%)	49(100%)	8(100%)	5(100%)	1(3%)	8(100%)	17(35%)	20(100%)	
Clinical Trial	158(38%)	96(48%)	0(0%)	28 (70%)	0(0%)	0(0%)	0(0%)	34(97%)	0(0%)	32(65%)	0(0%)	

* Kruskal-Wallis test

Abbreviations: BMI, body mass index; EE, ethinylestradiol, E2, estradiol; E4, estetrol; nAPCr, normalized activated protein C sensitivity ratio; SD, standard deviation

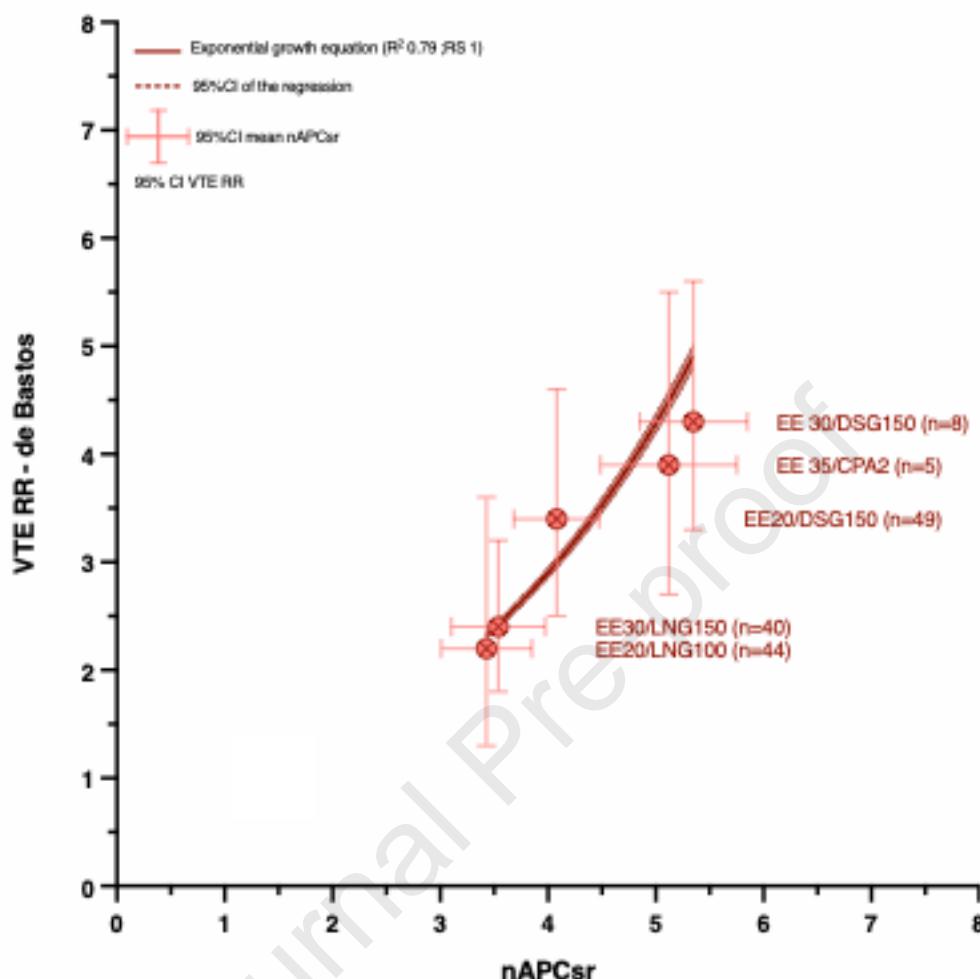
Table 2. Mean nAPCsr values (\pm SD), epidemiologically derived venous thromboembolism (VTE) risk (95%CI), and model-based estimated VTE risks (95%CI)

	COC non-users	EE 20 μ g - LNG 100 μ g	EE 30 μ g - LNG 150 μ g	EE 20 μ g - DSG 150 μ g	EE 30 μ g - DSG 150 μ g	EE 35 μ g - CPA 2mg	E4 15mg - DRSP 3mg	E2 1.5mg - NomAc 2.5mg	EE 20 μ g - DRSP 3mg	EE 30 μ g - DNG 2mg
nAPCsr										
Mean \pmSD	1.42 \pm 0.09	3.43 \pm 0.42	3.54 \pm 0.44	4.08 \pm 0.40	5.35 \pm 0.50	5.12 \pm 0.63	2.23 \pm 0.31	2.31 \pm 0.63	4.50 \pm 1.05	5.06 \pm 0.50
Epidemiologically derived VTE risk										
n (95%CI)	1 (1.3-3.6) ¹	2.2 (1.8-3.2) ¹	2.4 (2.5-4.6) ¹	3.4 (3.3-5.6) ¹	4.3 (2.7-5.5) ¹	-	1.6 (0.6-4.1) ^{*2}	4.84 (3.2-7.3) ³	3.4 (1.5-7.3) ^{*4}	
Model-based estimated VTE risk⁵										
n (95%CI)	1.06 (1.04-1.09)	2.32 (2.29-2.34)	2.42 (2.39-2.44)	2.98 (2.95-3.02)	4.89 (4.80-4.97)	4.47 (4.39-4.54)	1.45 (1.43-1.48)	1.50 (1.47-1.52)	3.43 (3.39-3.47)	4.36 (4.29-4.43)

¹ population-based risk from *de Bastos et al.* Cochrane Database Syst Rev. 2014; CD010813.² population-based risk from *Reed et al.* Eur J Contracept Reprod Health Care. 2021; 26: 439-46.³ population-based risk from *Lidegaard et al.* BMJ. 2011; 343⁴ population-based risk from *Dinger et al.* Frontiers in Women's Health. 2020; 5.⁵ Estimated using *in silico* model

Abbreviations: CI, confidence interval; COC, combined oral contraceptive; CPA, cyproterone acetate; DNG, dienogest; DSG, Desogestrel; EE, ethinylestradiol; E2, estradiol; E4, estetrol; LNG, levonorgestrel; nAPCsr, normalized activated protein C sensitivity ratio; NomAc, nomegestrol acetate; SD, standard deviation; VTE, venous thromboembolism

Supplementary Figure 1: Sensitivity analysis - model robustness after exclusion of the reference group



Abbreviations: *Cl*, confidence interval; *CPA*, cyproterone acetate; *DRSP*, drospirenone; *DSG*, desogestrel; *EE*, ethinylestradiol; *LNG*, levonorgestrel; *nAPCsr*, normalized activated protein C sensitivity ratio; *RR*, relative risk; *SD*, standard deviation; *VTE*, venous thromboembolism